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## Methylene blue inhibits replication of SARS-CoV-2 in vitro

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### ABSTRACT

In December 2019, a novel coronavirus, severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), causing coronavirus diseases 2019 (COVID-19) emerged in Wuhan, China. Currently there is no antiviral treatment recommended against SARS-CoV-2. Identifying effective antiviral drugs is urgently required. Methylene blue has already demonstrated in vitro antiviral activity in photodynamic therapy as well as antibacterial, antifungal and antiparasitic activities in non-photodynamic assays. In this study, non-photoactivated methylene blue showed in vitro activity at very low micromolar range with an EC<sub>50</sub> (median effective concentration) of  $0.30 \pm 0.03 \mu\text{M}$  and an EC<sub>90</sub> (90% effective concentration) of  $0.75 \pm 0.21 \mu\text{M}$  at a multiplicity of infection (MOI) of 0.25 against SARS-CoV-2 (strain IHUMI-3). The EC<sub>50</sub> and EC<sub>90</sub> values for methylene blue are lower than those obtained for hydroxychloroquine ( $1.5 \mu\text{M}$  and  $3.0 \mu\text{M}$ ) and azithromycin ( $20.1 \mu\text{M}$  and  $41.9 \mu\text{M}$ ). The ratios C<sub>max</sub>/EC<sub>50</sub> and C<sub>max</sub>/EC<sub>90</sub> in blood for methylene blue were estimated at 10.1 and 4.0, respectively, following oral administration and 33.3 and 13.3 following intravenous administration. Methylene blue EC<sub>50</sub> and EC<sub>90</sub> values are consistent with concentrations observed in human blood. We propose that methylene blue is a promising drug for treatment of COVID-19. In vivo evaluation in animal experimental models is now required to confirm its antiviral effects on SARS-CoV-2. The potential interest of methylene blue to treat COVID-19 needs to be confirmed by prospective comparative clinical studies.

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### 1. Introduction

In December 2019, a novel coronavirus, severe acute respiratory syndrome coronavirus (SARS-CoV-2), causing coronavirus diseases 2019 (COVID-19) emerged in Wuhan, China [1]. Despite containment measures, SARS-CoV-2 spread in Asia, Southern Europe, then in America and currently in Africa. Presently there is no antiviral treatment recommended against SARS-CoV-2. Different drugs or

combinations have been evaluated worldwide. Identifying effective low-cost antiviral drugs with limited side effects, affordable immediately, is urgently needed, especially for emerging countries.

Plasma products can transmit a wide range of pathogens by transfusion. Methylene blue, a synthesised thiazine dye, is known to be effective in photodynamic therapy against microbes and particularly viruses. Methylene blue is able to intercalate into viral nucleic acid when illuminated with visible light and prevents transmission of pathogens. Illumination of methylene blue inactivated Zika, yellow fever, dengue, chikungunya and Ebola viruses and Middle East respiratory syndrome coronavirus in plasma [2–5]. Methylene blue also demonstrates antimicrobial activities without photoactivation. Methylene blue inhibited in vitro colistin-resistant strains of *Acinetobacter baumannii*, *Mycobacterium ulcerans*, *Mycobacterium* spp. and *Candida albicans* [6–8]. Methylene blue was also effective in vivo against Buruli ulcer in experimental *M. ulcerans* infection in mice [7]. Additionally, methylene blue inacti-

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vated hepatitis C virus in transplant organ perfused with methylene blue [9]. The most studied effects of methylene blue are those on malaria.

In 1891, methylene blue was first used to effectively treat two patients with uncomplicated malaria [10]. In the 2010s, methylene blue showed effective in vitro activity in the nanomolar range against *Plasmodium falciparum* strains [11–14]. Methylene blue showed a protective effect against cerebral malaria in a murine model infected with *Plasmodium berghei* [15–17]. Methylene blue showed several benefits when used as a partner in triple combination with artemisinin-based combination therapy in uncomplicated falciparum malaria in children [18].

Taken together, these reports suggest that methylene blue may have antiviral effects against SARS-CoV-2. Therefore, in this study the activity of methylene blue was assessed in vitro against a clinically isolated SARS-CoV-2 strain and was compared with the activity of hydroxychloroquine and azithromycin, which have already been evaluated in vitro and in vivo in humans [19–22].

## 2. Materials and methods

### 2.1. Antimalarial drugs, virus and cells

Methylene blue (methylthioninium chloride; Proveblue®) was provided by Provepharm SAS (Marseille, France). Stocks solutions of hydroxychloroquine (Sigma, St Louis, MO, USA) and methylene blue were prepared in water, and azithromycin (Sigma) was prepared in methanol. All stock solutions were then diluted in Minimum Essential Medium (MEM) (Gibco, Thermo Fisher) to achieve seven final concentrations ranging from 0.1–100  $\mu\text{M}$ . A clinically isolated SARS-CoV-2 strain (IHUMI-3) [23] was maintained in production in Vero E6 cells (American Type Culture Collection ATCC® CRL-1586™) in MEM with 4% of fetal bovine serum (FBS) and 1% glutamine (complete medium).

### 2.2. Cytotoxicity assay

In vitro cell viability evaluation using the Vero E6 cell line was performed according to the method described by Mosmann with slight modifications [24]. Briefly,  $10^5$  cells in 200  $\mu\text{L}$  of complete medium were added to each well of 96-well plates and were incubated at 37 °C in a humidified 5%  $\text{CO}_2$  atmosphere. After 24 h of incubation, 25  $\mu\text{L}$  of complete medium and 25  $\mu\text{L}$  of each concentration of methylene blue, hydroxychloroquine or azithromycin were added and the plates were incubated for 48 h at 37 °C. After removal of the supernatant, 100  $\mu\text{L}$  of MTT [3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2H-tetrazolium bromide] (Sigma-Aldrich, France) solution (0.5 mg/mL in MEM without FBS) were then added to each well. Cells were incubated for 2 h at 37 °C. Following incubation, MTT solution was removed and 100  $\mu\text{L}$  of dimethyl sulfoxide (DMSO) was added to dissolve the formazan crystals. Plates were then shaken at 700 rpm for 10 min at 37 °C. The absorbance was measured at 570 nm using a Tecan Infinite F200 Microplate Reader. DMSO was used as a blank. The 50% cytotoxic concentration ( $\text{CC}_{50}$ ) was calculated with an inhibitory sigmoid  $E_{\text{max}}$  model, which estimated the  $\text{CC}_{50}$  through non-linear regression by using a standard function of the R software (ICEstimator v.1.2; <http://www.antimalarial-icestimator.net>). The  $\text{CC}_{50}$  value was the mean of six different experimentations.

### 2.3. Antiviral activity assay

Briefly, 96-well plates were prepared with  $5 \times 10^5$  cells/mL of Vero E6 cells (200  $\mu\text{L}$  per well) as previously described [20]. Methylene blue, hydroxychloroquine or azithromycin concentrations were added 4 h before infection. Vero E6 cells were in-

**Table 1**

$\text{EC}_{50}$  and  $\text{EC}_{90}$  values against SARS-CoV-2,  $\text{CC}_{50}$  and selectivity index (SI) for methylene blue, hydroxychloroquine and azithromycin

Drug	$\text{EC}_{50}$ ( $\mu\text{M}$ )	$\text{EC}_{90}$ ( $\mu\text{M}$ )	$\text{CC}_{50}$ ( $\mu\text{M}$ ) <sup>a</sup>	SI
Methylene blue	$0.30 \pm 0.03$	$0.75 \pm 0.21$	>100	>333
Hydroxychloroquine	$1.5 \pm 0.3$	$3.0 \pm 1.9$	$20.4 \pm 1.4$	13.6
Azithromycin	$20.1 \pm 4.5$	$41.9 \pm 18.0$	>100	>5

$\text{EC}_{50}$ , median effective concentration;  $\text{EC}_{90}$ , 90% effective concentration; SARS-CoV-2, severe acute respiratory syndrome coronavirus 2;  $\text{CC}_{50}$ , 50% cytotoxic concentration.

<sup>a</sup> In Vero E6 cells.

ected with SARS-CoV-2 strain IHUMI-3 at a multiplicity of infection (MOI) of 0.25. At 48 h post-infection, replication was estimated by RT-PCR using a SuperScript™ III Platinum™ One-Step Kit w/ROX (Invitrogen) after extraction with a BloExtract® SuperBall® Kit (Biosellal, Dardilly, France). The primers used have been described previously [25]. The  $\text{EC}_{50}$  (median effective concentration) and  $\text{EC}_{90}$  (90% effective concentration) were calculated with an inhibitory sigmoid  $E_{\text{max}}$  model, which estimated the  $\text{EC}_{50}$  and  $\text{EC}_{90}$  through non-linear regression using a standard function of the R software (ICEstimator v.1.2).  $\text{EC}_{50}$  and  $\text{EC}_{90}$  values were the mean of six different experimentations.

### 2.4. Data analysis and interpretation

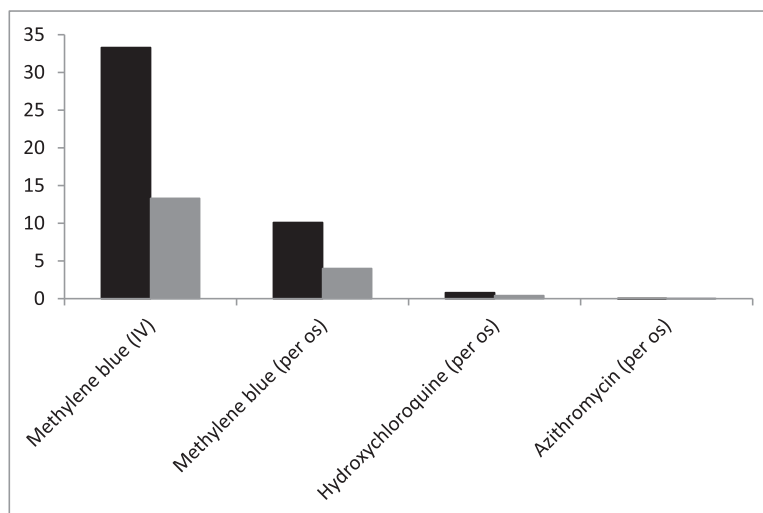
The selectivity index (SI) was estimated for each drug as the ratio of  $\text{CC}_{50}/\text{EC}_{50}$ . The expected maximum blood concentration ( $C_{\text{max}}$ ) was estimated from the literature for each drug at doses commonly administered in oral malaria treatment and for methylene blue at intravenous (i.v.) doses used for US Food and Drug Administration (FDA) and European Medicines Agency (EMA)-approved methemoglobinemia treatment. The ratios  $C_{\text{max}}/\text{EC}_{50}$  and  $C_{\text{max}}/\text{EC}_{90}$  were estimated to determine whether the effective concentration in plasma to cure SARS-CoV-2 is achievable in humans. If data on drug accumulation in the lung were available, the ratios  $C_{\text{lung}}/\text{EC}_{50}$  and  $C_{\text{lung}}/\text{EC}_{90}$  were calculated.

## 3. Results

The  $\text{CC}_{50}$ ,  $\text{EC}_{50}$ ,  $\text{EC}_{90}$  and SI for each drug are presented in Table 1. Methylene blue and hydroxychloroquine showed  $\text{EC}_{50}$  and  $\text{EC}_{90}$  values in the low micromolar range (Table 1). The  $\text{EC}_{50}$  and  $\text{EC}_{90}$  values for methylene blue were lower than those obtained for hydroxychloroquine and azithromycin. The ratios  $C_{\text{max}}/\text{EC}_{50}$  and  $C_{\text{max}}/\text{EC}_{90}$  in blood for methylene blue were estimated at 10.1 and 4.0, respectively, following oral administration and at 33.3 and 13.3 following i.v. administration (Fig. 1).

## 4. Discussion

Methylene blue showed in vitro activity at very low micromolar range with an  $\text{EC}_{50}$  of  $0.30 \pm 0.03 \mu\text{M}$  and an  $\text{EC}_{90}$  of  $0.75 \pm 0.21 \mu\text{M}$  at a MOI of 0.25 (SI > 333) (Table 1). The  $\text{EC}_{50}$  and  $\text{EC}_{90}$  values for methylene blue are lower than those obtained for hydroxychloroquine and azithromycin. Azithromycin demonstrated low in vitro efficacy against SARS-CoV-2 when used alone but potentiated the effects of hydroxychloroquine in combination [20]. Oral uptake of 325 mg of methylene blue led to a  $C_{\text{max}}$  in blood of 0.97  $\mu\text{g}/\text{mL}$  (~3  $\mu\text{M}$ ) and an elimination half-life ( $t_{1/2}$ ) of 14.9 h [26]. A methylene blue dose of 2 mg/kg i.v. showed a  $C_{\text{max}}$  of 2.917  $\mu\text{g}/\text{mL}$  (~10  $\mu\text{M}$ ) [27]. The ratios  $C_{\text{max}}/\text{EC}_{50}$  and  $C_{\text{max}}/\text{EC}_{90}$  for methylene blue were estimated at 10.1 and 4.0 for the oral route and 33.3 and 13.3 for the i.v. route, respectively. Methylene blue  $\text{EC}_{50}$  and  $\text{EC}_{90}$  values are consistent with concentrations observed in human blood. Approximately 3–5% of methylene blue per gram of lung was found



**Fig. 1.** Bar chart displaying the ratios  $C_{max}/EC_{50}$  (black) and  $C_{max}/EC_{90}$  (grey) for methylene blue, hydroxychloroquine and azithromycin for in vitro activity against SARS-CoV-2 (severe acute respiratory syndrome coronavirus 2).  $C_{max}$ , maximum concentration in blood;  $EC_{50}$ , median effective concentration;  $EC_{90}$ , 90% effective concentration.

after i.v. methylene blue injection but the methylene blue concentration decreased rapidly below 0.1% after 10 h [28]. In comparison, oral uptake of 400 mg of hydroxychloroquine led to a  $C_{max}$  of 1.22  $\mu\text{M}$  [29]. Hydroxychloroquine accumulated 30 times more in the lungs than in blood [30]. The azithromycin  $C_{max}$  ranged from 0.18–0.4  $\mu\text{g}/\text{mL}$  of blood ( $\sim 0.22$ – $0.51 \mu\text{M}$ ) after the last dose of oral administration of 500 mg once daily for 3 days or after a single dose of 500 mg [31–33]. These doses led to a  $C_{max}$  in the lung ranging from 8–9  $\mu\text{g}/\text{g}$  ( $\sim 10$ – $12 \mu\text{M}$ ) [31,32]. The  $C_{max}$  expected in the lung was below the  $EC_{50}$  and  $EC_{90}$ . However, due to potentiation of the antiviral effects when azithromycin is combined with hydroxychloroquine, azithromycin can be used in vitro at lower concentrations (5  $\mu\text{M}$  and 10  $\mu\text{M}$ ) [20]. These concentrations are compatible with expected concentrations in the lungs.

Methylene blue showed low cytotoxicity in vitro against Vero E6 cells with  $CC_{50} > 100 \mu\text{M}$ . The SI as a ratio of  $CC_{50}/EC_{50}$  was estimated to be  $>333$ . The present  $CC_{50}$  of hydroxychloroquine with an SI of  $\sim 13$  against Vero E6 cells was higher than previously reported  $CC_{50}$  values, ranging from  $>50 \mu\text{M}$  to  $250 \mu\text{M}$  against Vero E6 cells [19,34] or  $>500 \mu\text{M}$  in *Felis catus* whole fetus-4 cells [35]. Azithromycin also showed low cytotoxicity against Vero E6 cells with  $CC_{50} > 100 \mu\text{M}$  and  $SI > 5$ . The  $CC_{50}$  for azithromycin was consistent with previous data ( $>130 \mu\text{M}$ ) [34]. Methylene blue showed low cytotoxicity but predominantly the higher SI.

Although methylene blue is on the list of drugs potentially dangerous for patients with glucose-6-phosphate dehydrogenase (G6PD) deficiency, no association between methylene blue and severe haemolysis has been detected after oral administration [36]. Additionally, the i.v. route for methylene blue has been granted a marketing authorisation in Europe in 2011 and in the USA in 2016 for the treatment of acquired methemoglobinemia based upon a confirmed positive benefit/risk ratio in this pathology.

## 5. Conclusion

Methylene blue showed high in vitro antiviral effective activity against SARS-CoV-2 with an  $IC_{50}$  (0.3  $\mu\text{M}$ ) and  $IC_{90}$  (0.75  $\mu\text{M}$ ) compatible with oral uptake and i.v. administration. This in vitro activity is higher than those obtained with drugs that have been evaluated in clinical trials worldwide such as hydroxychloroquine (1.5  $\mu\text{M}$ ), azithromycin (20.1  $\mu\text{M}$ ), remdesivir (23  $\mu\text{M}$ ), lopinavir (26.6  $\mu\text{M}$ ) or ritonavir ( $>100 \mu\text{M}$ ) [37]. We propose that methylene blue is a promising drug for the treatment of COVID-19. In

vivo evaluation in animal experimental models is now required to confirm its antiviral effects against SARS-CoV-2. The potential interest of methylene blue to treat COVID-19 needs to be confirmed by prospective comparative clinical studies.

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## References

- [1] Wu F, Zhao S, Yu B, Chen YM, Wang W, Song ZG, et al. A new coronavirus associated with human respiratory disease in China. *Nature* 2020;579:365–9.
- [2] Fryk JJ, Marks DC, Hobson-Peters J, Prow NA, Watterson D, Hall RA, et al. Dengue and chikungunya viruses in plasma are effectively inactivated after treatment with methylene blue and visible light. *Transfusion* 2016;56:2278–85.
- [3] Faddy HM, Fryk JJ, Hal RA, Young PR, Reichenberg S, Tolksdorf F, et al. Inactivation of yellow fever virus in plasma after treatment with methylene blue and visible light and in platelet concentrates following treatment with ultraviolet C light. *Transfusion* 2019;59:2223–7.
- [4] Wang Y, Ren K, Liao X, Luo G, Kumthip K, Leetrakool N, et al. Inactivation of Zika virus in plasma and derivatives by four different methods. *J Med Virol* 2019;91:2059–65.
- [5] Eickmann M, Gravemann U, Handke W, Tolksdorf F, Reichenberg S, Müller TH, et al. Inactivation of Ebola virus and Middle East respiratory syndrome coronavirus in platelet concentrates and plasma by ultraviolet C light and methylene blue plus visible light, respectively. *Transfusion* 2018;58:2202–7.
- [6] Gazel D, Tatman Otkun M, Akçali A. In vitro activity of methylene blue and eosin methylene blue agar on colistin-resistant *A. baumannii*: an experimental study. *J Med Microbiol* 2019;68:1607–13.
- [7] Tian RBD, Asmar S, Napez C, Lépidi H, Drancourt M. Effectiveness of purified methylene blue in an experimental model of *Mycobacterium ulcerans* infection. *Int J Antimicrob Agents* 2017;49:290–5.

- [8] Pal R, Ansari MA, Saibabu V, Das S, Fatima Z, Hameed S. Nonphotodynamic roles of methylene blue: display of distinct antimycobacterial and anticandidal mode of actions. *J Pathog* 2018;2018:3759704.
- [9] Helfritz FA, Bojkova D, Wanders V, Kuklinski N, Westhaus S, von Horn C, et al. Methylene blue treatment of grafts during cold ischemia time reduces the risk of hepatitis C virus transmission. *J Infect Dis* 2018;218:1711–21.
- [10] Guttman P, Ehrlich P. Ueber die Wirkung des Methylenblau bei Malaria [About the effect of methylene blue in malaria]. *Berl Klin Wochenschr* 1891;28:953–6.
- [11] Pascual A, Henry M, Briolant S, Charras S, Baret E, Amalvict R, et al. In vitro activity of Proveblue (methylene blue) on *Plasmodium falciparum* strains resistant to standard antimalarial drugs. *Antimicrob Agents Chemother* 2011;55:2472–4.
- [12] Fall B, Camara C, Fall M, Nakoulima A, Dionne P, Diatta B, et al. *Plasmodium falciparum* susceptibility to standard and potential anti-malarial drugs in Dakar, Senegal, during the 2013–2014 malaria season. *Malar J* 2015;14:60.
- [13] Fall B, Madamet M, Diawara S, Briolant S, Wade KA, Lo G, et al. Ex vivo activity of Proveblue, a methylene blue, against field isolates of *Plasmodium falciparum* in Dakar, Senegal from 2013 to 2015. *Int J Antimicrob Agents* 2017;50:155–8.
- [14] Gendrot M, Madamet M, Mosnier J, Fonta I, Amalvict R, Benoit N, et al. Baseline and multinormal distribution of ex vivo susceptibilities of *Plasmodium falciparum* to methylene blue in Africa, 2013–18. *J Antimicrob Chemother* 2020;75:2141–8.
- [15] Dormoi J, Pradines B. Dose responses of Proveblue methylene blue in an experimental murine cerebral malaria model. *Antimicrob Agents Chemother* 2013;57:4080–1.
- [16] Dormoi J, Briolant S, Desgrouas C, Pradines B. Efficacy of Proveblue (methylene blue) in an experimental cerebral malaria murine model. *Antimicrob Agents Chemother* 2013;57:3412–14.
- [17] Dormoi J, Briolant S, Desgrouas C, Pradines B. Impact of methylene blue and atorvastatin combination therapy on the apparition of cerebral malaria in a murine model. *Malar J* 2013;12:127.
- [18] Mendes Jorge M, Ouermi L, Meissner P, Compaoré G, Coulibaly B, Nebie E, et al. Safety and efficacy of artesunate–amodiaquine combined with either methylene blue or primaquine in children with falciparum malaria in Burkina Faso: a randomized controlled trial. *PLoS One* 2019;14:e0222993.
- [19] Liu J, Cao R, Xu M, Wang X, Zhang H, Hu H, et al. Hydroxychloroquine, a less toxic derivative of chloroquine, is effective in inhibiting SARS-CoV-2 infection in vitro. *Cell Discov* 2020;6:16.
- [20] Andreani J, Le Bideau M, Dufлот I, Jardt P, Rolland C, Boxberger M, et al. In vitro testing of hydroxychloroquine and azithromycin on SARS-CoV-2 shows synergistic effect. *Microb Pathog* 2020;145:104228.
- [21] Gautret P, Lagier JC, Parola P, Hoang VT, Meddeb L, Sevestre J, et al. Clinical and microbiological effect of a combination of hydroxychloroquine and azithromycin in 80 COVID-19 patients with at least a six-day follow up: an observational study. *Travel Med Infect Dis* 2020;34:101663.
- [22] Million M, Lagier JC, Gautret P, Colson P, Fournier PE, Amrane S, et al. Early treatment of 1061 COVID-19 patients with hydroxychloroquine and azithromycin, Marseille, France. *Travel Med Infect Dis* 2020;35:101738.
- [23] Gautret P, Lagier JC, Parola P, Hoang VT, Meddeb L, Mailhe M, et al. Hydroxychloroquine and azithromycin as a treatment of COVID-19: results of an open-label non-randomized clinical trial. *Int J Antimicrob Agents* 2020;56:105949.
- [24] Mosmann T. Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. *J Immunol Methods* 1983;65:55–63.
- [25] Amrane S, Tissot-Dupont H, Doudier B, Eldin C, Hocquart M, Mailhe M, et al. Rapid viral diagnosis and ambulatory management of suspected COVID-19 cases presenting at the infection diseases referral hospital in Marseille, France, January 31st to March 1st, 2020: a respiratory virus snapshot. *Travel Med Infect Dis* 2020;36:101632.
- [26] Anh CX, Chavchich M, Birrell GW, van Breda K, Travers T, Rowcliffe K, et al. Pharmacokinetics and ex vivo antimalarial activity of artesunate–amodiaquine plus methylene blue in healthy volunteers. *Antimicrob Agents Chemother* 2020;64 e01441-19.
- [27] Center for Drug Evaluation and Research Clinical pharmacology and biopharmaceutics review(s): application number 204630Orig1s000; October 2020. [https://www.accessdata.fda.gov/drugsatfda\\_docs/nda/2016/204630Orig1s000ClinPharmR.pdf](https://www.accessdata.fda.gov/drugsatfda_docs/nda/2016/204630Orig1s000ClinPharmR.pdf) [accessed 16 October 2020].
- [28] Link EM, Costa DC, Lui D, Ell PJ, Lower PJ, Spittle MF. Targeting disseminated melanoma with radiolabelled methylene blue. *Acta Oncol* 1996;35:331–41.
- [29] Rainsford KD, Parke AL, Clifford-Rashotte M, Kean WF. Therapy and pharmacological properties of hydroxychloroquine and chloroquine in the treatment of systemic lupus erythematosus, rheumatoid arthritis and related diseases. *Inflammopharmacology* 2015;23:231–69.
- [30] Chhonker YS, Sleightholm RL, Li J, Oupicky D, Murry DJ. Simultaneous quantification of hydroxychloroquine and its metabolites in mouse blood and tissues using LC-ESI-MS/MS: an application for pharmacokinetic studies. *J Chromatogr B Analyt Technol Biomed Life Sci* 2018;1072:320–7.
- [31] Danesi R, Lupetti A, Barbara C, Ghelardi E, Chella A, Malizia T, et al. Comparative distribution of azithromycin in lung tissue of patient given oral daily doses of 500 and 1000 mg. *J Antimicrob Chemother* 2003;51:939–45.
- [32] Lucchi M, Damle B, Fang A, de Caprariis PJ, Mussi A, Sanchez SP, et al. Pharmacokinetics of azithromycin in serum, bronchial washings, alveolar macrophages and lung tissue following a single oral dose of extended or immediate release formulations of azithromycin. *J Antimicrob Chemother* 2008;61:884–91.
- [33] Davidson RJ. In vitro activity and pharmacodynamic/pharmacokinetic parameters of clarithromycin and azithromycin: why they matter in the treatment of respiratory tract infections. *Infect Drug Resist* 2019;12:585–96.
- [34] Madrid PB, Panchal RG, Warren TK, Shurleff AC, Endsley AN, Green CE, et al. Evaluation of Ebola virus inhibitors for drug repurposing. *ACS Infect Dis* 2015;1:317–26.
- [35] Takano T, Satoh K, Doki T, Tanabe T, Hohdatsu T. Antiviral effects of hydroxychloroquine and type I interferon on in vitro fatal feline coronavirus infection. *Viruses* 2020;12:576.
- [36] Lu G, Nagbanshi M, Goldau N, Mendes Jorge M, Meissner P, Jahn A, et al. Efficacy and safety of methylene blue in the treatment of malaria: a systemic review. *BMC Med* 2018;16:59.
- [37] Choy KT, Wong AYL, Kaewpreedee P, Sia SF, Chen D, Hui KPY, et al. Remdesivir, lopinavir, emetine, and homoharringtonine inhibits SARS-CoV-2 replication in vitro. *Antiviral Res* 2020;178:104786.